

# Journal of Ecology and The Natural Environment

April 2018 ISSN 2006-9847 DOI: 10.5897/JENE www.academicjournals.org



# **ABOUT JENE**

The Journal of Ecology and the Natural Environment (JENE) (ISSN 2006-9847) is published Monthly (one volume per year) by Academic Journals.

**Journal of Ecology and the Natural Environment (JENE)** provides rapid publication (monthly) of articles in all areas of the subject such as biogeochemical cycles, conservation, paleoecology, plant ecology etc.

The Journal welcomes the submission of manuscripts that meet the general criteria of significance and scientific excellence. Papers will be published shortly after acceptance. All articles published in JENE are peer-reviewed.

### **Contact Us**

Editorial Office:	jene@academicjournals.org
Help Desk:	helpdesk@academicjournals.org
Website:	http://www.academicjournals.org/journal/JENE
Submit manuscript online	http://ms.academicjournals.me/

# **Editors**

### Dr. Abd El-Latif Hesham

Genetics Department, Faculty of Agriculture, Assiut University, Assiut 71516, Egypt

**Dr. Ahmed Bybordi** East Azarbaijan Research Centre for Agriculture and Natural Resources, Tabriz, Iran

# Dr. Sunil Kumar

Natural Resource Ecology Laboratory, Colorado State University 1499 Campus Delivery, A204 NESB, Fort Collins, Colorado-80526, USA

# Prof. Gianfranco Rizzo

University of Palermo Dipartimeno DREAM – Viale delle Scienze - Building 9. 90128 Palermo, Italy

# Dr. Bahman Jabbarian Amiri

Kiel University, Germany, Ökologie-Zentrum der CAU Abt. Hydrologie und Wasserwirtschaft Olhausen Straße, 75 Kiel, Germany

# Dr. Bikramjit Sinha

National Institute of Science Technology and Development Studies, Pusa Gate, Dr. KS Krishnan Marg, New Delhi 110012, India

# Prof. Gianfranco Rizzo

University of Palermo Dipartimeno DREAM – Viale delle Scienze - Building 9. 90128 Palermo, Italy

# **Associate Editors**

### Dr. Marko Sabovljevic

Dept. Plant Ecology, Faculty of Biology, University of Belgrade Takovska 43, 11000 Belgrade, Serbia

# Dr. Sime-Ngando Télesphore

CNRS LMGE, UMR 6023, Université Blaise Pascal, 63177, Aubière Cedex France

# Dr. Bernd Schierwater

ITZ, Ecology and Evolution, TiHo Hannover Büenteweg 17d, 30559 Hannover, Germany

# Dr. Bhattacharyya Pranab

North-East Institute of Science & Technology Medicinal, Aromatic & Economic Plant Division, North-East Institute of Science & Technology, Jorhat-785006, Assam, India

# **Prof. Marian Petre**

University of Pitesti, Faculty of Sciences 1 Targul din Vale Street, Pitesti, 110040, Arges County, Romania.

# Prof. R.C. Sihag

CCS Haryana Agricultural University Department of Zoology & Aquaculture, Hisar-125004, India

# Prof. Kasim Tatic

School of Economics and Business, University of Sarajevo Trg oslobodjenja 1, 71000 SARAJEVO, Bosnia and Herzegovina

# Dr. Zuo-Fu Xiang

Central South University of Forestry & Technology, 498 Shaoshan Nanlu, Changsha, Hunan, China.

#### Dr. Zuo-Fu Xiang

Central South University of Forestry & Technology, 498 Shaoshan Nanlu, Changsha, Hunan, China.

#### Dr.Pankaj Sah

Higher College of Technology, Muscat, Department of Applied Sciences, (Applied Biology) Higher College of Technology, Al-Khuwair, PO Box 74, PC 133, Muscat (Sultanate of Oman)

#### Dr. Arti Prasad

Mohan Lal Sukhadia University, Udaipur,Rajasthan,india. 123,Vidya Nagar,Hiran Magri, Sector-4,Udaipur,Rajasthan, India

# **Editorial Board**

#### Parviz Tarikhi

Mahdasht Satellite Receiving Station (Postal): No. 80, 14th Street, Saadat Abad Avenue, Tehran 1997994313, Iran

#### **Bharath Prithiviraj**

Post Doctoral Research Associate Knight Lab, Dept. of Chemistry & Biochemistry University of Colorado at Boulder USA

#### Dr. Melissa Nursey-Bray

Australian Maritime College, Tasmania, Australia

#### Parvez Rana

Department of Forestry and Environmental Science Shahjalal University of Science and Technology Bangladesh

#### Mirza Hasanuzzaman

Faculty of Agriculture, Sher-e-Bangla Agricultural University Sher-e-Bangla Nagar, Dhaka-1207, Bangladesh

#### Dr. Giri Kattel

Murray Darling Freshwater Research Centre, La Trobe University 471 Benetook Avenue, Mildura, Victoria 3500, Australia

#### Dr. M. Rufus Kitto

Faculty of Marine Science-Obhur station, King Abdulaziz University, Jeddah 21589, Saudi Arabia

#### Dr. Özge Zencir

Kemah Vocational Training School, Erzincan University, Kemah, Erzincan, Turkey.

#### Dr. Sahadev Sharma

Laboratory of Ecology and Systematics, Graduate School of Engineering and Science, University of the Ryukyus,Senbaru 59, Nishihara, Okinawa-903-0213 Japan

#### Dr. Hasan Kalyoncu

University of Süleyman Demirel, Faculty of Art and Science, Departmant of Biology, 32100 Isparta/Turkey

#### Hammad Khan

Department of Zoology and Fisheries, University of Agriculture, Faisalaad,Pakistan

#### Mirza Hasanuzzaman

Faculty of Agriculture, Sher-e-Bangla Agricultural University Sher-e-Bangla Nagar, Dhaka-1207, Bangladesh

#### Abdurrahman Dundar

Siirt University, Science and Arts Faculty, Department of Biology, 56000, Siirt, Turkey

#### Meire Cristina Nogueira de Andrade

College of Agronomic Sciences, São Paulo State University, Brazil.

#### Imran Ahmad Dar

Dept. of Industries and Earth Sciences, The Tamil University, Ocean and Atmospheric Sciences & Technology Cell, (A Unit of Ministry of Earth Sciences, Govt. of India).

#### S. Jayakumar

Department of Ecology and Environmental Sciences, School of Life Sciences, Pondicherry University, Puducherry - 605 014, India

#### Umer Farooq

University of Veterinary & Animal Sciences Lahore, Pakistan

# Journal of Ecology and the Natural Environment

Table of Contents: Volume 10 Number 3, April 2018

# **ARTICLE**

Mycorrhizal status of some indigenous tree species in the Takamanda rainforest, South West Region, Cameroon Eneke Esoeyang Tambe Bechem, Njoh Roland Ndah and Egbe Enow Andrew

41

# academicJournals

Vol. 10(3), pp. 41-52, April 2018 DOI: 10.5897/JENE2018.0684 Article Number: 7A6BD9656929 ISSN 2006-9847 Copyright © 2018 Author(s) retain the copyright of this article http://www.academicjournals.org/JENE

Journal of Ecology and The Natural Environment

Full Length Research Paper

# Mycorrhizal status of some indigenous tree species in the Takamanda rainforest, South West Region, Cameroon

Eneke Esoeyang Tambe Bechem<sup>1\*</sup>, Njoh Roland Ndah<sup>1,2</sup> and Egbe Enow Andrew<sup>1</sup>

<sup>1</sup>Department of Botany and Plant Physiology, University of Buea, P. O. Box 63 Buea, Cameroon. <sup>2</sup>Forests, Resources and People, Limbe P. O. Box 111 Limbe, Cameroon.

Received 14 February, 2018; Accepted 28 March, 2018

A survey was carried out to determine the type of mycorrhizal association formed by trees within the different habitat types of the disturbed and undisturbed sites of the Takamanda rainforest. Forty-eight tree species of commercial and cultural importance were selected from the two sites for this study. Root samples were collected from a total of 327 individual trees belonging to the 48 species; they were cleared, stained and examined microscopically for mycorrhizal colonization and type. All the forty-eight species examined harbored one or more mycorrhizal structures, which ranged from arbuscules, intercellular hyphae, intracellular hyphae, vesicles, and Hartig net. Thirty-nine species formed exclusively arbuscular mycorrhiza (81.25%), two species; Uapaca guineensis and Angylocalyx oligophyllus formed ectomycorrhiza only (4.17%), while seven species Afzelia bipindensis, Baphia nitida, Anglylocalyx pynaertii, Cieba pentandra, Cylicodiscus gabunensis, Pterocarpus soyauxii and Terminalia ivorensis formed both ecto- and arbuscular mycorrhiza (14.58%). In both forest sites and habitat types, arbuscular mycorrhiza was the most represented among the tree species. In the undisturbed site and in the plain 68% of tree species sampled formed arbuscular mycorrhiza, 12% formed ectomycorrhiza, 16% formed dual mycorrhiza and 4% were non-mycorrhiza. On ridge top, 81.8% of the tree species formed arbuscular mycorrhiza, 13.6% formed ectomycorrhiza with 4.6% being dual mycorrhiza. On hilly slopes, 82.8% of the tree species formed arbuscular mycorrhiza, 13.8% formed ectomycorrhiza and 3.5% were dual mycorrhiza. In the disturbed site, 100% of the tree species sampled on the plain, formed arbuscular mycorrhiza. On the ridge top, 73.3% of the tree species sampled formed arbuscular mycorrhiza, 13.3% formed ectomycorrhiza and 13.3% were non mycorrhizal. On hilly slopes, 83.3% formed arbuscular mycorrhiza, 8.3% formed ectomycorrhiza and 8.3% were non-mycorrhizal. Mycorrhizas are important factors in Takamanda and must be taken into consideration, when designing management strategies for this forest.

Key words: Arbuscular mycorrhiza, ectomycorrhiza, Takamanda forest.

# INTRODUCTION

The management and conservation of forest biomes is a recognized priority on a global scale. Understanding the rhizosphere and the mycorrhizal associations in particular, is important for the proper management of

forest ecosystems. Mycorrhizal relationship (mutualistic associations between specialized Basidio-, Asco-, and Glomeromycetous fungi and roots of higher plants) constitute the most efficient nutrient uptake facilitators particularly in nutrient deficient soils of tropical regions (Ike-Izundu, 2001; Onguene and Kuyper, 2001; Kernagham, 2005). Mycorrhizal symbioses are ubiquitous in terrestrial ecosystems and have been shown to exert significant influence on recruitment, species composition, richness and productivity of plant communities (Vander Heijden et al., 1998; Moyersoen and Fitter, 1999; Terwilliger and Pastor, 1999; Weber et al., 2005). However, the degree of this influence in the rainforest of Takamanda is yet to be unraveled.

A number of different types of mycorrhizas exist in nature and can be identified by the hyphal structures they form. Arbuscular mycorrhizas (AM), sometimes referred to as endomycorrhizas, are formed predominantly by the fungi of the Glomeromycota (Schubler et al., 2001). Their name is derived from structures they form within cells of plant root, arbuscules. Arbuscules are finely-branched structures that are formed within a cell and serve as a major metabolic exchange site between the plant and the fungus. Vesicles are formed by some species of AM fungi; they are sac-like structures, emerging from hyphae, which serve as storage organs for lipids (Smith and Read, 2008). AM are formed by a wide range of plant species, belonging to different life forms. AM colonization has no visible effect on root morphology (Brundrett, 2009). Ectomycorrhizas (EM), on the other hand, form an outer sheath (mantle) around the root, an internal, intercellular network of hyphae (Hartig net) and rhizomorphs. EM fungi have a visible effect on root morphology. Root tip branching often becomes dichotomous or irregular. The root tip also tends to swell and the mantle may colour the area of colonization (Hawley and Dames, 2004). It is widely accepted that most tropical tree species form mostly AM and limited species form EM associations (Onguene and Kuyper, 2000; Skinner, 2001; Bechem and Alexander, 2012; Bechem et al., 2014). EM associations have been observed in selected genera in the humid tropics, the Caesalpiniaceae, primarily from Fabaceae. Gnetaceae, Euphorbiaceae, Myrtaceae, Nyctaginaceae, Araucariaceae, and Polygonaceae (Alexander and Hogberg, 1986; Alexander, 1989; Henkel et al., 2002; Wang and Qiu, 2006). Substantive research has been done on EM associations in the Dipterocarpaceae of Asia (Alexander, 1989; Alexander et al., 1992; Lee et al., 1997), Caesalpiniaceae and the Uapacaceae of Africa (Hogberg and Piearce, 1986; Newbery et al., 1988; Torti and Coley, 1999).

Nevertheless, there are many species for which mycorrhizal associations have to be examined (Skinner, 2001; Alexander and Lee, 2005). Mycorrhizal status, as well as the influence of mycorrhizal mutualism-parasitism

continuum on rainforest diversity is poorly known compared to the wealth of botanical and ecological studies in these ecosystems (Alexander and Lee, 2005; Bechem and Alexander, 2012). The Takamanda rainforest is popularly known for being the home to the endangered cross river gorilla, besides this, it also harbors a diverse variety of plant species whose mycorrhiza status is still to be determined in this habitat. The determination of the mycorrhiza status of plants in this habitat will help in the elaboration of management strategies for the forest. This study was therefore undertaken to examine the mycorrhizal status of some indigenous tree species of the Takamanda rainforest Cameroon.

#### MATERIALS AND METHODS

#### Study area and tree selection

The Takamanda rainforest is situated on the Southern corner of the South West Region of Cameroon between 05°59' to 06°21'N and 09°11 to 09°30'E (Figure 1). This study was carried out in the undisturbed and disturbed sites of the Takamanda rainforest. The undisturbed site is located within the Takamanda National Park, thus access is restricted. The disturbed site is located outside the park and is very accessible to members of the community. The distance separating the two sites is about 3.5 km. The landscape of the area is undulating (ridge top, hilly slope and plain) in both undisturbed and disturbed forests sites (Figure 2). The undulating nature of the forest sites resulted to a variety of habitat types (plain, ridge top and hilly slope).

For the mycorrhizal screening, 48 tree species of economic and cultural importance were selected. The tree species listed in Table 1, comprise of 25 commercialized timber species (T) and 23 tree species that provide non-timber forest products (N). Some of the tree species such as *Pterocarpus* soyauxii, Annikia chlorantha and Cylicodiscus gabunensis offered multiple uses to the communities (Zapfack et al., 2001; Ndah et al., 2013). Out of the 48 tree species selected, 14 species occurred in both undisturbed and disturbed forest sites, 9 species solely in disturbed site and 25 species only in the undisturbed forest. In the undisturbed forest, the roots of 25 tree species were sampled in the plain, 22 were sampled in the ridge top and 29 in the hilly slope, while in the disturbed site, 9 species were sampled in the plain, 15 species in ridge top and 12 in hilly slope.

The 48 selected mature tree species for mycorrhizal evaluation were identified in the field, tagged and root samples were collected in the undisturbed and disturbed forest sites of the Takamanda rainforest. At least three individuals per species per site were sampled in order to ascertain the consistency of mycorrhizal type. In the sample collection process, soil at the base of each tree was dug-up using a hand fork and roots were followed from the base of the tree to the feeder roots to ensure accuracy of identification. Roots were collected from the four cardinal directions around each tree. Root samples were collected between June to July 2015 in both undisturbed and disturbed forest sites. Approximately 15 to 20 g (fresh weight) of fine root material was collected for each species in sites where they existed; soil was washed-off carefully

<sup>\*</sup>Corresponding author. E-mail: tamenekeso@yahoo.co.uk.

Author(s) agree that this article remains permanently open access under the terms of the <u>Creative Commons Attribution</u> <u>License 4.0 International License</u>

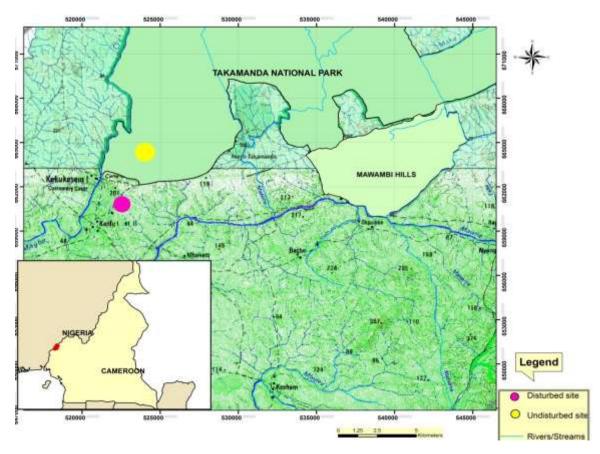


Figure 1. Map of Akwaya showing the study sites (CIFOR landscape map).

with fresh water and the samples were stored in labeled vials containing 50% alcohol (Brundrett et al., 1996) for onward transportation to the life sciences laboratory at the University of Buea. Whilst in the laboratory, samples were stored in the refrigerator at 4°C and were processed within 72 h from collection.

# Processing and examination of root samples for mycorrhizal structures

For detailed microscopic examination, representative root samples were washed carefully with tap water to remove alcohol; soil particles and adhering organic matter were removed with the aid of a fine brush, forceps and under a dissecting microscope. A subsample of the roots was then cleared and stained using the procedure described by Brundrett et al., (1996). Root samples were cleared by immersing them in 10% KOH for 2 days. They were then rinsed several times in tap water to remove the clearing solution (Brundrett et al., 1996; Onguene and Kuyper 2001; Hawley and Dames, 2004). They were stained in a 0.05% trypan blue in lactoglycerol for 2 to 3 days. Stained roots were rinsed in tap water and cut with a razor blade into approximately 2 cm long pieces. For each species, five slides were prepared, such that 25 root fragments of up to 50 cm were placed on a glass slide and gently squashed under a cover slip to observe the presence and type of mycorrhiza structures under a compound microscope (Olympus BX 41). Arbuscular mycorrhiza was characterized by the presence of intracellular hyphae, hyphal and arbusculate coils and arbuscules, while ectomycorrhiza were recorded by the presence of intercellular hyphae, mantle and Hartig net (Brundrett, 2008; Onguene, 2000).

In the categorization of the type of mycorrhiza that occurred in a plant, a plant species was scored as EM when ectomycorrhiza structures only were observed in sampled individuals, it was AM when arbuscular mycorrhiza structures only were observed, it was AM\_EM when both ecto- and arbuscular mycorrhiza structures were observed in the roots of plants of the same species. Photographs of mycorrhizal structures observed were captured with an Olympus DP 20 camera attached to the microscope.

#### Data analysis

Information gathered here on the occurrence of mycorrhiza types on the different species was compared to data from other forest habitats in order to draw conclusions on mycorrhiza habits of each of the species studied. The effect of habitat types and sites on mycorrhizal colonization of tree species was also evaluated using a two way analysis of variance.

#### RESULTS

# Mycorrhizal status of tree species in the disturbed and undisturbed rainforest

All 48 tree species in both undisturbed and disturbed forests formed mycorrhizas (Tables 2 and 3). Out of the 48 tree species in the two sites, tree species formed

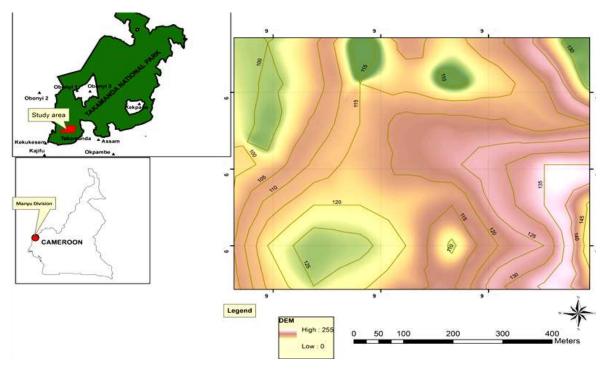


Figure 2. Study site, showing the undulating nature of the terrain (CIFOR landscape map).

arbuscular mycorrhiza (81.25%), only two species (*Uapaca guineensis* and *Angylocalyx oligophyllus*) formed ectomycorrhiza (4.16%) (Tables 2, 3) and seven of them (*Afzelia bipindensis*, *Baphia nitida*, *Anglylocalyx pynaertii*, *Ceiba pentandra*, *Cylicodiscus gabunensis*, *Pterocarpus soyauxii* and *Terminalia ivorensis*) formed both ecto- and arbuscular mycorrhizas (14.58%) in roots of the same individuals or on different individuals of the same species. This was characterized by the presence of intracellular hyphae, intercellular hyphae; arbuscules and Hartig net (Tables 2 and 3; Figure 3a to d).

In both forest sites and in all habitat types, AM was the most prevalent mycorrhiza formed by the tree species in this study. At the undisturbed site, AM was recorded in 17 (68%) species in plain, 18 (81.8%) on ridge top and 24 (82.8%) on hilly slope while in the disturbed site AM was observed in 9 (100%) species in the plain, 11 (73.3%) on ridge top and 10 (83.3%) species in hilly slope (Tables 2 and 3). This was closely followed by EM at undisturbed site with 3 (12%) species in plain, 3 (13.6%) species in ridge top and 4 (13.8%) species on hilly slope. In the disturbed site EM was observed in 2 (13.3%) of species on ridge top and in 1 (8.3%) in the hilly slope (Table 3). Generally, dual mycorrhizal (AM EM) colonization was lowest in both sites and habitats. At the undisturbed site AM\_EM was seen in 4 (16%) species in the plain, 1 (4.6%) species in the ridge top and in 1 (3.5%) species on hilly slope (Table 2). Dual mycorrhiza colonization was observed in 1 (8.3%) species sampled in the hilly slope in the disturbed forest. From the results of the two way ANOVA, there was no significant (P>0.05) difference in mycorrhiza occurrence for the species in both forest sites and in the different habitat types. In the undisturbed forest, all plants sampled in the different habitat types formed mycorrhizas except in the plain, where 1 out of the 25 plant species was non-mycorrhizal, giving a 96% mycorrhiza occurrence. Similarly, in the disturbed site, all plants sampled in the different habitats harbored mycorrhizal structures, resulting to a 100% mycorrhiza occurrence, except in the ridge top where 2 out of the 15 plants sampled were non-mycorrhizal with 86.7% occurrence.

Some inconsistencies in mycorrhiza types were observed for some tree species at both sites and in the different habitat types (Table 4). Afzelia bipindensis formed dual mycorrhizas and harbored both AM and EM structures on roots of the same plant as well as from different plants collected from plain and hilly slopes. Angylocalyx pynaertii and Pterocarpus soyauxii also formed dual mycorrhizal associations with individual root samples collected from the plain (low land) showing both AM and EM structures (Table 4). Five of the seven with dual mycorrhiza associations species were Fabaceae (Table 4). Figures 3a to d shows some of the mycorrhizal structures observed on sampled plants. This included intracellular hyphae, arbuscules and intercellular hyphae.

# DISCUSSION

All tree species studied were mycorrhizal. We observed

Family	Species	Category
Anacardiaceae	Lannea welwitschii (Hiern) Engl.	Т
Anacardiaceae	Pseudospondias microcarpa (A. Rich.) Engl.	Т
<b>A</b>	Annickia chlorantha (Oliv.) Setten & Maas.	Ν
Annonaceae	Anonidium mannii (Oliv.)	Ν
Anon/100000	Funtumia elastica (Preuss) Stapf	т
Apocynaceae	Rauvolfia vomitoria Afzel	Ν
Bombacaceae	Ceiba pentandra (L.) Gaertn.	т
Burseraceae	Dacryodes edulis (G. Don) H. J. Lam	Ν
Cecropiaceae	Musanga cecropioides R. Br.	Ν
Clusiaceae	Garcinia kola Heckel	Ν
Combretaceae	Terminalia ivorensis A. Chev.	т
Dichapetalaceae	Tapura africana Engl.	т
Ebenaceae	Diospyros preussii Gürke	Ν
	Mallotus oppositifolius (Geisel.) MüllArg.	Ν
Euphorbiaceae	<i>Plagiostyles africana</i> (Müll.Arg.) Prain	Т
Luphorbiaceae	Ricinodendron heudelottii (Baill.) Pierre	Ν
	Uapaca guineensis MüllArg.	Ν
	Afzelia bipindensis Harms.	т
	<i>Albizia zygia</i> (DC.) J.F. Macbr.	Т
	Amphimas pterocarpoides Harms	Т
	Angylocalyx oligophyllus (Baker) Baker f.	Ν
	Angylocalyx pynaertii De Wild.	N
	Anthonotha macrophylla P.Beauv.	N
Fabaceae	Baphia nitida Lodd.	N
	Cylicodiscus gabunensis Harms.	T
	Distemonanthus benthamianus Baill.	Т
	Hylodendron gabunense Taub.	T
	Parkia bicolor A. Chev.	T T
	Piptadeniastrum africanum (Hook.f.) Brenan Calpocalyx dinklagei Harms.	N
	Pterocarpus soyauxii Taub.	Т
Flacourtiaceae	Homalium logistylum Alex.	т
Irvingiaceae	Irvingia gabonensis (Aubrey-Lecomte ex O. Rorke). Baill.	N
Meliaceae	Lovoa trichilioides Harms.	т
Moraceae	Ficus exasperate Vahl.	N
MUIALEAE	Milicia excelsa (Welw.) C.C.Berg	Т

 Table 1. Tree species selected for evaluation of mycorrhizal status in the Takamanda rainforest.

Myristicaceae	Pycnanthus angolensis (Welw.) Warb. Staudtia stipitata Warb.	T T
Olacaceae	Strombosia grandifolia Hook. f.	Ν
Polygalaceae	<i>Carpolobia lutea</i> (G. Don)	Ν
Pandaceae	Microdesmis puberula Hook. f. ex Planch.	Ν
Passifloraceae	Barteria fistulosa Mast.	Ν
Rubiaceae	Mitragyna ciliata (Aubrev. & Pellegr)	т
Sapindaceae	Blighia welwitschii (Hiern) Radlk.	Т
Sterculiaceae	Cola millenii K. Schum Eribroma oblogum Mast. Sterculia tragacantha Lindl.	N T T
Violaceae	Rinorea dentata (P Beauv.) Kuntze	Ν

Table 1. Contd.

T: Commercialized Timber species and N: Non timber forest products.

that majority of the tree species examined harbored AM fungal structures. The high AM status could probably be attributed to the production of large quantities of viable spores and large spore sizes containing substantial energy during adverse conditions, thus AM inoculum might rapidly build up in the system than EM. Our observations were similar to those of other surveys in Africa, which showed most or all of the woody species being mycorrhizal with a majority forming AM. Hawley and Dames (2004) assessed mycorrhizal status of 15 indigenous tree species in South Africa and all were AM. In a similar survey carried out in Tanzania, out of the forty-seven indigenous trees and shrubs species investigated, forty of them were reported to be AM and the remaining seven species EM (Hogberg, 1982). Onguene and Kuyper (2001) assessed the mycorrhizal status of 100 tree species in a rainforest of South Cameroon. All the species were mycorrhizal, with 26 species forming EM and the remaining species being AM. Newbery et al. (1988) in Korup National Park, Cameroon also reported that, out of the 56 species investigated, 55 turned out to be mycorrhizal with only one species, Wareneckea memecyloides, which is а Meslastomataceae being non-mycorrhizal. Bechem et al., (2014) also reported a similar finding in a lowland rainforest of Cameroon with a 92.06% of species studied being AM and 3.97% forming EM. The results in this study further confirm the fact that trees forming AM dominate tropical rainforests. Arbuscular mycorrhiza fungi thrive in P deficient soils in the tropics. They have the ability to access insoluble P, making it available to their host plant, in addition to other benefits (Smith and Read, 2008). AM fungi demonstrate little specificity with respect to host plant, thus AM plants dominate tropical forests.

Mycorrhizas structures were absent in some individuals of Cylicodiscus gabunensis, Lovoa trichiliodes, Staudtia stiapitata, Albizia zygia, Barteria fistulosa and Ceiba pentandra in some habitat types in this study. However, some of these species showed mycorrhizas association in other habitats. Birhance et al. (2010) mentioned that environmental characteristics can influence colonization therefore our observations may be due to the differences in characteristics of the different habitat types, since elevation changed from plain to hilly slopes to ridge tops. Uapacca guineensis was the only species which formed EM in both forest sites. This is in conformity with findings of Onguene and Kuyper (2001) and Bechem et al. (2014) who also observed EM structures on roots of these species. This could be attributed to species specificity to particular types of mycorrhiza. Other species of this genus Uapaca have been reported to show dual mycorrhizal status (Onguene and Kuyper, 2001). Seven tree species formed dual mycorrhizal associations; but four of the species Afzelia bipindensis, Baphia nitida, Cylicodiscus gabunensis and Pterocarpus soyauxii all belonging to the Fabaceae, harbored both AM and EM structures in the same root sample. This observation was similar to that of studies carried out in Central western Guyana (McGuire et al., 2008); South Cameroon (Onguene and Kuyper, 2001) Korup National Park

<b>Femil</b> u	Species		F	Plain		Ridge tops				Hilly slopes			
Family	Species	AM	EM	AM_EM	NM	AM	EM	AM_EM	NM	AM	EM	AM_EM	NM
Area and a second	Lannea welwischii	-	-	-	-	-	-	-	-	Х	0	0	0
Anacardiaceae	Pseudospondias microcarpa	-	-	-	-	-	-	-	-	Х	0	0	0
Annonaceae	Annickia chlorantha	Х	0	0	0	-	-	-	-	х	0	0	0
Annonaceae	Anonidium mannii	-	-	-	-	-	-	-	-	Х	0	0	0
Apocynaceae	Funtumia elastica	Х	0	0	0	Х	0	0	0	х	0	0	0
Apocynaceae	Rauvolfia vomitoria	Х	0	0	0	Х	0	0	0	Х	0	0	0
Bombacaceae	Ceiba pentandra	0	Х	0	0	Х	0	0	0	-	-	-	-
Cecropiaceae	Musanga cecropioides	Х	0	0	0	-	-	-	-	Х	0	0	0
Combretaceae	Terminalia ivorensis	Х	0	0	0	х	Х	0	0	0	х		0
Dichapetalaceae	Tapura africana	Х	0	0	0	х	0	0	0	х	0	0	0
Ebenaceae	Diosoyros preussi	Х	0	0	0	-	-	-	-	-	-	-	-
Funkarkinggan	Ricinodendron heudeloti	-	-	-	-	Х	0	0	0	х	0	0	0
Euphorbiaceae	Uapaca guineensis	0	Х	0	0	-	-	-	-	0	Х	0	0
	Afzelia bipindensis	0	Х	х	0	Х	0	0	0	х	0	х	0
	Albizia zygia	-	-	-	-	-	-	-	-	0	Х	0	0
Fabaceae	Amphimas pterocarpoides	-	-	-	-	Х	0	0	0	-	-	-	-
ravalleae	Angylocalyx oligophyllus	0	Х	0	0	-	-	-	-	-	-	-	-
	Angylocalyx pynaertii	0	Х	Х	0	0	Х	0	0	-	-	-	-
	Anthonotha macrophylla	-	-	-	-	0	Х	0	0	-	-	-	-

Table 2. Mycorrhizal status of tree species at different habitat types in the undisturbed Takamanda rainforest.

AM: Arbuscular mycorrhiza, EM: Ectomycorrhiza, AM\_EM: Both ecto and arbuscular mycorrhiza, NM: Non mycorrhiza, X: Presence of mycorrhizal type(s), and 0: Absence of mycorrhizal type - = not sampled in habitat.

(Bechem et al., 2014). *Terminalia ivorensis* was AM in plain and EM in ridge top; *Ceiba pentandra* was EM in plain and AM in ridge top. These inconsistencies could be attributed to

environmental variability and availability of inocula in these micro habitat types. Mycorrhizal status was generally consistent for most tree species in the same habitats and forest sites. Similar consistent observations have been reported by Nandakwang et al. (2008); Evelin et al. (2009); Birhane et al. (2010). Tree root colonization consisted of intercellular and intracellular hyphae,

### Table 2. Contd.

Family	Species		I	Plain		Ridge tops				Hilly slopes			
Family		AM	EM	AM-EM	NM	AM	EM	AM-EM	NM	AM	EM	AM-EM	NM
	Calpocalyx dinklagei	-	-	-	-	0	Х	0	0	0	Х	0	0
	Cylicodiscus gabunensis	Х	0	Х	Х	-	-	-	-	-	-	-	-
	Distemonanthus benthamianus	Х	0	0	0	Х	0	0	0	Х	0	0	0
<b>F</b> -h	Hylodendron gabunense	Х	0	0	0	Х	0	0	0	Х	0	0	0
Fabaceae	Parkia bicolor	-	-	-	-	-	-	-	-	Х	0	0	0
	Piptademastrum africana	Х	0	0	0	-	-	-	-	Х	0	0	0
	Pterocarpus soyauxii	Х	Х	Х	0	Х	0	0	0	Х	0	0	0
Flacourtiaceae	Homalium logistylum	-	-	-	-	-	-	-	-	х	0	0	0
Icacinaceae	Lasianthera africana	-	-	-	-	-	-	-	-	х	0	0	0
Irvingiaceae	Irvingia gabonensis	Х	0	0	0	-	-	-	-	-	-	-	-
Meliaceae	Lovoa trichliodes	0	0	0	Х	-	-	-	-	х	0	0	0
Myristicaceae	Pycnanthus angolensis	Х	0	0	0	Х	0	0	0	х	0	0	0
Mynsicaceae	Staudtia stipitata	-	-	-	-	Х	0	0	Х	-	-	-	-
Olacaceae	Strombosia grandifolia	Х	0	0	0	-	-	-	-	-	-	-	-
Pandaceae	Microdesmis puberula	Х	0	0	0	Х	0	0	0	х	0	0	0
Polygalaceae	Carpolobia lutea	-	-	-	-	х	0	0	0	х	0	0	0
Rubiaceae	Mitragyna ciliata	-	-	-	-	-	-	-	-	Х	0	0	0
Sapindaceae	Blighia welwitschii	-	-	-	-	х	0	0	0	Х	0	0	0
Charaulianas -	Eribroma oblogum	Х	0	0	0	Х	0	0	0	х	0	0	0
Sterculiaceae	Sterculia tragacantha	Х	0	0	0	Х	0	0	0	Х	0	0	0
Violaceae	Rinorea dentata	Х	0	0	0	х	0	0	0	-	-	-	-

AM: Arbuscular mycorrhiza, EM: Ectomycorrhiza, AM\_EM: Both ecto and arbuscular mycorrhiza, NM: Non mycorrhiza, X: presence of mycorrhizal type (s) and 0: Absence of mycorrhizal type - =not sampled in habitat.

<b>Femily</b>	Species	Plain				Ridge tops				Hilly slopes			
Family		AM	EM	AM_EM	NM	AM	EM	AM_EM	NM	AM	EM	AM_EM	NM
Anacardiaceae	Lannea welwischii	-	-	-	-	Х	0	0	0	-	-	-	-
Bombacaceae	Cieba pentandra	-	-	-	-	Х	0	0	0	Х	0	0	0
Burseraceae	Dacryodes edulis	Х	0	0	0	-	-	-	-	-	-	-	-
Cecropiaceae	Musanga cecropioides	-	-	-	-	Х	0	0	0	Х	0	0	0
Clusiaceae	Garcinia Kola	Х	0	0	0	-	-	-	-	-	-	-	-
Combretaceae	Terminalia ivorensis	-	-	-	-	-	-	-	-	0	Х	0	0
	Mallotus oppositifolius	-	-	-	-	Х	0	0	0	-	-	-	-
Euphorbiaceae	Ricinodendron heudeloti	Х	0	0	0	Х	0	0	0	Х	0	0	0
	Uapaca guineensis	-	-	-	-	0	Х	0	0	0	Х	0	0
	Albizia zygia	Х	0	0	х	Х	0	0	0	х	0	0	0
	Amphimas pterocarpoides	-	-	-	-	0	0	0	Х	-	-	-	-
Fabaceae	Baphia nitida	Х	0	0	0	Х	0	0	0	Х	0	Х	0
	Distemonanthus benthamianus	-	-	-	-	Х	0	0	0	Х	0	0	0
	Pterocarpus soyauxii	Х	0	0	0	0	Х	0	0	-	-	-	-
Irvingiaceae	Irvingia gabonensis	Х	0	0	0	-	-	-	-	Х	0	0	0
Managan	Ficus exaspirata	-	-	-	-	-	-	-	-	х	0	0	0
Moraceae	Milicia excelsa	Х	0	0	0	-	-	-	-	Х	0	0	0
Myristicaceae	Pycnanthus angolensis	Х	0	0	0	Х	0	0	0	-	-	-	-
Starouliagoago	Cola millenii	-	-	-	-	-	-	-	-	х	0	0	0
Sterculiaceae	Eribroma Oblogum	-	-	-	-	Х	0	0	0	-	-	-	-
Pandaceae	Microdesmis puberula	-	-	-	-	х	0	0	0	-	-	-	-

 Table 3. Mycorrhizal status of tree species at different habitats in the disturbed Takamanda rainforest.

AM: Arbuscular mycorrhiza, EM: Ectomycorrhiza, AM\_EM: both ecto and arbuscular mycorrhiza, NM: Non mycorrhiza, X: presence of mycorrhizal type (s), and 0: Absence of mycorrhizal type - =not sampled in habitat.

Family	Species	Plain	Ridge tops	Hilly slopes
Bombacaceae	Cieba pentandra	EM	AM	-
Combretaceae	Terminalia ivorensis	AM	AM, EM	EM
	Afzelia bipindensis	EM, AM_EM	AM	AM, AM_EM
	Angylocalyx pynaertii	EM, AM_EM	EM	-
Fabaceae	Baphia nitida	AM	AM	AM_EM
	Cylicodiscus gabunensis	AM	AM_EM	-
	Pterocarpus soyauxii	AM, EM, AM_EM	AM	AM

Table 4. Inconsistency in mycorrhizal types in the different habitats of the Takamanda Rainforest.

AM\_EM: AM and EM Structures on roots of same individual; AM, EM: Structures on roots of different individuals of same species; - tree species not sampled in the habitat hilly slope.

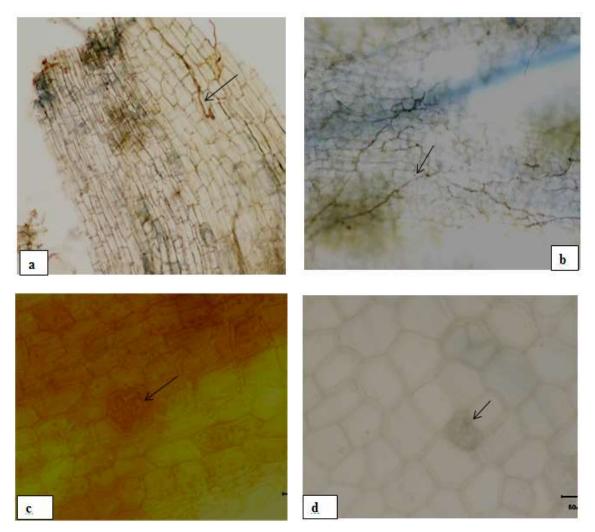


Figure 3. Sections of roots showing intercellular hyphae (i and ii), arbuscules (iii) and endophytes (iv).

arbuscules, and Hartig net. The dominance of intracellular and intercellular hypha structures in the different habitats and forest types indicated the presence of favourable environmental conditions which promoted the growth of the mycelia (Brundrett et al., 1996; Hawley and Dames, 2004; Becerra et al., 2007; Nandakwang et al., 2008). Birhane et al. (2010); Becerra et al. (2007); Brundrett (2009) an d Moyersoen et al. (1998) mentioned that environmental conditions may influence the colonization and development of mycorrhizal structures. Studies have shown that anthropogenic activities resulting to the removal and replacement of species has an effect on the fungal diversity of the soil (Paillet et al., 2010). Management measures for the different forest sites must therefore be designed such that below ground diversity is protected. In the study reported here more of the plant species sampled occurred in the undisturbed site. It would be interesting to know whether this above ground diversity in the undisturbed site is proportional to the below ground diversity.

#### Conclusions

This study provides the first account of mycorrhizal status of tree species in the Takamanda rainforest. AM was the most prevalent mycorrhizal type in the Takamanda rainforest. Dual colonization of tree species in this rainforest was reported with species belonging to the Fabaceae and Combretaceae. Mycorrhizal colonization did not show great differences across habitat types and the forest sites of the Takamanda rainforest. Proper management of the forest ecosystem will improve mycorrhizal colonization, thus enhance nutrient uptake, circulation, resistance to diseases and would encourage the growth of tree species. Management strategies for the Takamanda National Park must be designed in such a way as to promote mycorrhiza formation and guarantee the continuous existence of the inoculum.

#### CONFLICT OF INTERESTS

The authors have not declared any conflict of interests.

#### ACKNOWLEDGMENTS

The authors are thankful to the management of FOREP (Forests, Resources and People) for providing partial funding for this work. The authors are also grateful to the management of the Infectious Disease Laboratory, University of Buea for storage of plant roots and viewing of mycorrhiza structures. Authors appreciate grant in the form of equipment from IDEA WILD which were used for data collection and processing and finally to the chiefs and communities of the Takamanda rainforest who accepted this research work in their area. The authors are thankful to the anonymous reviewers, whose comments have helped to improve the quality of the manuscript.

#### REFERENCES

Alexander IJ, Hogberg P (1986). Ectomycorrhizas of tropical angiospermous trees. New Phytologist 102:541-549.

- Alexander IJ (1989). Mycorrhizas in tropical forests: In: Proctor, J (Ed.). Mineral nutrients in Tropical Forest and Savanna Ecosystems. Blackwell Scientific Publications, Oxford, pp. 169-188.
- Alexander IJ, Lee SS (2005). Mycorrhizas and ecosystem processes in tropical rain forest: implications for diversity. In: Burslem DFRP, Pinard MA, Hartley SE (eds) Biotic interactions in the tropics: their role in the maintenance of species diversity. Cambridge University Press, Cambridge, UK, pp. 165-203.
- Alexander I, Ahmad N, Lee SS (1992). The role of mycorrhizae in the regeneration of some Malaysian forest trees. Philosophical Trans. Royal Soc. B, 335:379-388.
- Becerra A, Cabello M, Chiarini F (2007). Arbuscular mycorrhizal colonization of vascular plant from the Yungas forest Argentina. Annals For. Sci. 64:765-772.
- Bechem, EET, Alexander IJ (2012). Mycorrhiza status of *Gnetum* spp.in Cameroon: evaluating diversity with a view to ameliorating domestication efforts. Mycorrhiza 22:99-108.
- Bechem, EET, Chuyong B, Fon, TB (2014). A survey of mycorrhizal colonization in the 50-ha Korup Forest Dynamic Plot in Cameroon. Am. J. Plant Sci. 5:1403-1415.
- Birhane E, Kuyper WT, Sterck JF, Bongers F (2010). Arbuscular mycorrhizal associations with *Boswellia papyrifera* (Frankincence tree) dominated by dry deciduous wood land of Northern Ethiopia. J. For. Ecol. Manag. 260:2160-2169.
- Brundrett M (2009). Mycorrhizas in natural ecosystem. J. Adv. Ecol. Res. 21:171-313.
- Brundrett MC (2008). Ectomycorrhizas. In: Mycorrhizal Associations: The Web Resource. Version 2.0 Data accessed. (mycorrhizas.info).
- Brundrett M, Bougher N, Dell B, Grove T, Malajezuk N (1996). Working with mycorrhizas in forestry and agriculture ACIAR Monographs 32. http://aciar.gov.au/publication/mn032
- Evelin H, Rupan K, Bhoopander G (2009). Arbuscular mycorrhizal fungi in alleviation of salt stress: a review. Annals Bot. 1:1-18.
- Habte M (2000). Mycorrhiza fungi and plant nutrition: In: J. A. Silva and R. Uchida, (eds.). Plant Nutrition Management in Hawaii's Soils, Approaches for Tropical and subtropical Agricultural College of tropical and Agricultural and Human Resources University of Hawaii at Manoa pp. 127-131.
- Hawley LG, Dames FJ (2004). Mycorrhizal status of indigenous tree species in the forest biomes of the Eastern Cape, South Africa. South Afr. J. Sci. 100:633-637.
- Henkel TW, Terborgh JT, Vilgalys R. (2002). Ectomycorrhizal fungi and their leguminous hosts in the Pakaraima Mountains of Guyana. Mycol. Res. 106:515-531.
- Hogberg P. (1982). Mycorrhizal associations in some woodlands and forest trees and shrubs in Tanzania *New Phytologist* 92:407-415
- Hogberg P, Piearce GD. (1986). Mycorrhizas in Zambian trees in relation to host taxonomy, vegetation type and successional patterns. J. Ecol. 74:775-785.
- Ike-Izundu NE (2007). Interaction between Arbuscular Mycorrhizal fungi and soil Microbial Populations in the Rhizosphere. Msc. Thesis Rhodes University pp. 25-32.
- Kernaghan G (2005). Mycorrhizal diversity: Cause and effect? Pedobiologia 49:511-520.
- Lee LS, Alexander IJ, Watling R (1997). Ectomycorrhizas and putative ectomycorrhizal fungi of *Shorea leprosula* Miq (Dipterocarpaceae). Mycorrhiza 7:63-81.
- McGuire KL, Henkel, TW, Granzow I, Villa G, Edmund F, Andrew C (2008). Dual mycorrhizal colonization of forest-dominating tropical trees and the mycorrhizal status of non-dominant tree and liana species. Mycorrhiza 18(4):217-222.
- Moyersoen B, Alexander IJ, Fitter AH (1998). Phosphorus nutrition of ectomycorrhizal and arbuscular mycorhizal tree seedlings from a lowland tropical rain forest in Korup National Park. Cameroon J. Trop. Ecol. 14:47-61.
- Moyersoen B, Fitter AH (1999). Presence of arbuscular mycorrhizas in typically ectomychorrhizas host species from Cameroon and New Zealand. Mycorrhiza 8:247-253.
- Nandakwang P, Elliot S, Youpensuk S, Dell B, Teaumroong N, Lumyong S (2008). Arbuscular mycorrhizal of indigenous tree species used to restore seasonally dry tropical forest of Northern Thailand. Res. J. Microbiol. 3(2):51-61.

- Ndah NR, Egbe AE, Bechem E, Asaha S, Yengo T, Chia EL, Eyenieh MN (2013). Ethnobotanical Study of Commonly used Medicinal Plants of the Takamanda Rainforest South West, Cameroon. Afr. J. Plant Sci. 7(1):21-34.
- Newbery DM, Alexander IJ, Thomas DW, Gartlan JS (1988). Ectomycorrhizal rainforest legumes and soil phosphorus in Korup National Park, Cameroon. New Phytologist 109:433-450.
- Newbery MD, Chuyong BG, Zimmermann L, Praz C (2006). Seedling survival and growth of three ectomycorrhizal Caesalpineaceous tree species in a central African rainforest. J. Trop. Ecol. 22:499-511.
- Onguene NA (2000). Diversity and dynamics of mycorrhizal associations in tropical rainforest with different disturbance regime in south Cameroon. Tropenbos-Cameroon Series 3, 167p.
- Onguene, NA, Kuyper TW (2001). Mycorrhizal associations in rainforest of South Cameroon For. Ecol. Manag. 140:277-287.
- Paillet Y, Bergès L, Hjältén J, Ódor P, Avon C, Bernhardt-Römermann MA, Bijlsma RJ, De Bruyn LU, Fuhr M, Grandin UL, Kanka R 2010. Biodiversity differences between managed and unmanaged forests: meta-analysis of species richness in Europe. Conservation Biol. 24(1):101-12.
- Schubler A, Schwarzott D, Walker C (2001). A new fungal phylum, the Glomeromycota: phylogeny and evolution. Mycol. Res. 105(12):1413-1421.

- Smith SE, Read DJ (2008). *Mycorrhizal Symbiosis*. 3<sup>rd</sup> edn. Academic Press.
- Skinner A (2001). A mycorrhizal survey of indigenous plant species within the Featherstone Kloof and DassieKrantz area on the Grahamstown Commonage, Eastern Cape, South Africa. B.Sc. (Hons) Thesis, Rhodes University, Grahamstown 89p.
- Terwilliger J, Pastor J (1999). Small mammals, ectomycorrhizae, and conifer succession in beaver meadows. Oikos 85:83-95.
- Torti SD, Coley PD (1999). Tropical monodominance: a preliminary test of the ectomycorrhizal hypothesis. Biotropic 3:220-228.
- Treseder KK (2004). A meta-analysis of mycorrhizal responses to nitrogen, phosphorus, and atmospheric  $CO_2$  in field studies. New Phytol. 164:347-355.
- Van-der Heijden MGA, Klironomos JM, Ursie M, Moutoglis P, StreitwolfEngel R, Boller T, Wiemken A, Sanders IR (1998). Mycorrhizal fungal diversity determines plant biodiversity, ecosystem variability and productivity. Nat. 396(6706):69.
- Wang B, Qiu YL (2006). Phylogenetic distribution and evolution of mycorrhizas in land plants. Mycorrhiza 72:299-363.
- Weber A, Karst J, Gilbert B, Kimmins JP (2005). *Thujaplicata* exclusion in ectomycorrhiza-dominated forests: testing the role of inoculum potential of arbuscular mycorrhizalfungi. Oecologia 143:148-156.

# **Related Journals:**



African Journal of **Microbiology Res** arch

icsandSequenceAndy





www.academicjournals.org